

## FULL PAPER

Clerodendrumol, A New Triterpenoid from *Clerodendrum yaundense* GÜRKE (Lamiaceae)by Maurice D. Awouafack<sup>\*a)</sup>, Simayijiang Aimaiti<sup>b)</sup>, Pierre Tane<sup>a)</sup>, and Hiroyuki Morita<sup>\*b)</sup><sup>a)</sup> Laboratory of Natural Products Chemistry, Department of Chemistry, Faculty of Science, University of Dschang, P.O. Box 67, Dschang, Cameroon

(phone: +237 69986 1628; fax: +237 2434 51735; e-mail: ducret.awouafack@univ-dschang.org; amauduc2@yahoo.com)

<sup>b)</sup> Institute of Natural Medicine, University of Toyama, 2630-Sugitani, Toyama 930-0194, Japan

(phone: +81 76 434 7625; fax: +81 76 434 5059; e-mail: hmorita@inm.u-toyama.ac.jp)

A new lupane-type triterpene named clerodendrumol (**1**), *i.e.*, (16 $\alpha$ )-lupa-12,20(29)-dien-16-ol, together with five known compounds, octacos-(5Z,9Z)-dienoic acid (**3**), tetracosanoic acid (**4**), genkwanin (**5**), (12S)-hydroxyoctadeca-(9Z,13E,15E)-trienoic acid (**6**), and (9S)-hydroxyoctadeca-(10E,12E)-dienoic acid (**7**), were isolated from the twigs of *Clerodendrum yaundense*. *O*-Acetylclerodendrumol (**2**), a derivative of **1**, was prepared by a standard acetylation procedure applied to compound **1**. The structure of the new compound was elucidated using spectroscopic analysis (NMR and MS) data, and chemical conversions (acetylation and deacetylation). The human breast cancer cell line (MDA-MB-231) was sensitive to all test samples, and clerodendrumol (**1**) as well as its derivative **2** had good activity ( $IC_{50}$  values 7.5 and 13.2  $\mu$ M). Genkwanin (**5**) exhibited strong activity on HeLa cell line with  $IC_{50}$  7.8  $\mu$ M.

**Introduction.** – *Clerodendrum*, a member of the family Lamiaceae, is a very large and diverse genus with approximately 600 species distributed in Asia, Australia, America, and Africa [1]. Plant species of this genus are small trees, shrubs and herbs, and have several traditional medicine uses in Asia and Africa [2][3]. Previous phytochemical investigations on some species of the genus *Clerodendrum* reported the isolation of phenols, flavonoids, terpenoids, steroids, and cyanogenic glycosides [1][2][4–6].

*Clerodendrum yaundense* is a small tree of 2 to 4 m high with white flowers found in the western region of Cameroon where traditional healers used the leaves for the treatment of jaundice and infectious diseases. To the best of our knowledge, no phytochemical and biological studies have been reported on this plant species.

In part of our continuing search for bioactive constituents from plant species of *Clerodendrum* [4], we report the isolation and structure elucidation of a new triterpenoid (**1**) and a derivative **2** thereof, together with five known compounds (**3–7**) from the twigs of *C. yaundense*. The structure of the new compound was established based on spectroscopic analysis (NMR and MS), and chemical conversions. The antiproliferative activity of the crude extract, **1–3**, and **5** is also reported.

**Results and Discussion.** – The MeOH crude extract of the twigs of *Clerodendrum yaundense* was subjected to silica gel medium pressure liquid column chromatography (MPLC). Then, several attempts were made to separate the mixture by column chromatography (*Sephadex LH-20*, normal-phase and reversed-phase  $SiO_2$ ), but only chemical

resolution by acetylation/deacetylation was successful to afford clerodendrumol (**1**) and *O*-acetylclerodendrumol (**2**) along with five known compounds, octacos-(5Z,9Z)-dienoic acid (**3**) [7], tetracosanoic acid (**4**) [8], genkwanin (**5**) [9], (12S)-hydroxyoctadeca-(9Z,13E,15E)-trienoic acid (**6**) [10], and (9S)-hydroxyoctadeca-(10E,12E)-dienoic acid (**7**) [11] (*Fig. 1*). Compounds **1** and **3** were initially isolated as a mixture. The acetylation reaction of the mixture of **1** and **3** gave compounds **2** and **3**, while deacetylation of **2** furnished **1**. The structures of the known compounds were identified by comparison of their spectroscopic data with those reported in the literature.

Clerodendrumol (**1**) was obtained as white powder and the IR spectrum had prominent absorption bands at 3438 and 1645  $cm^{-1}$ , characteristic for a OH group and of non-conjugated C=C bonds. The HR-FAB-MS gave a *pseudo*-molecular ion peak ( $[M+H]^+$ ) at  $m/z$  425.3781 corresponding to the molecular formula  $C_{30}H_{48}O$ , consistent with seven degrees of unsaturation. The  $^1H$ -NMR spectrum (*Table I*) of **1** had signals ( $\delta(H)$  5.31 (br. *d*,  $J=4.6$ ), 4.74 (br. *s*), and 4.67 (br. *s*)) of a vinylic and of exocyclic  $CH_2$  group H-atoms. A signal due to a H-atom of an O-bearing CH group was also observed at  $\delta(H)$  3.40–3.33 (*m*). The  $^1H$ -NMR spectrum exhibited further signals of three H-atoms each, for seven *singlets* between  $\delta(H)$  0.72 and 1.58, corresponding to one Me group attached to a vinylic C-atom, and six tertiary Me groups. The  $^{13}C$ -NMR spectrum (*Table I*) exhibited a total number of 30 C-atoms including seven C-atoms, six CH, ten  $CH_2$ , and seven Me. Important C-atom signals were observed at  $\delta(C)$  148.2 (C(20)), 142.2 (C(13)), 121.6 (C(12)), and 112.2 (C(29)) characteristic for a lupane-type triterpenoid [12][13]. The  $^{13}C$ -NMR spec-

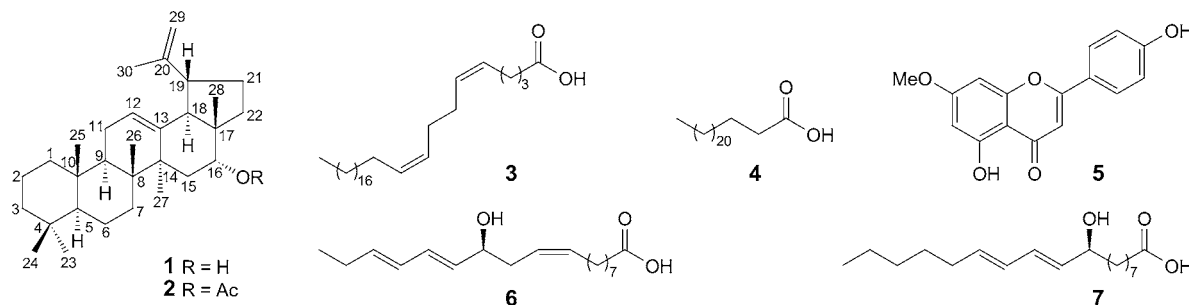


Fig. 1. Chemical structures of compounds 1–7

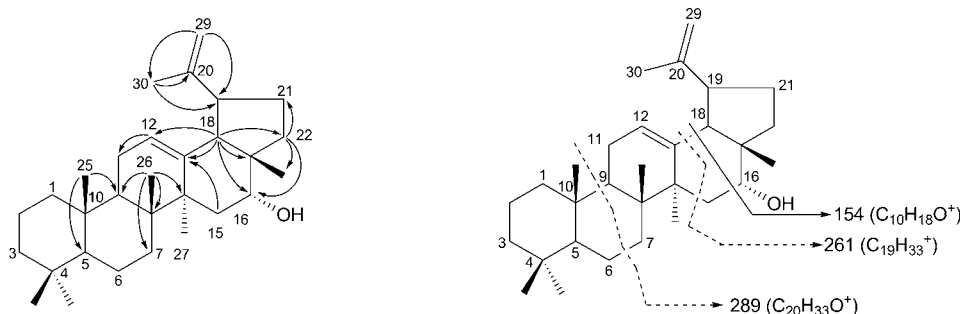


Fig. 2. Key HMBCs (H → C) and MS fragments of clerodendrumol (1)

trum had an additional and relevant C-atom signal at  $\delta(C)$  71.8 assignable to C(16). The HMBCs (Fig. 2) between H-atom signals at  $\delta(H)$  2.21 (H–C(18)), 1.83–1.81 ( $H_a$ –C(22)) and C-atom signal at  $\delta(C)$  71.8 (C(16)) was indicative for the OH group to be attached to C(16). The position of the OH group at C(16) was further supported by ion fragments observed in the mass spectrum (Fig. 2) at  $m/z$  154 ( $[M+H-275]^+$ ,  $C_{10}H_{18}O^+$ ; C(13)–C(18) and C(15)–C(16) cleavage), 195 ( $[M+H-428]^+$ ,  $C_{14}H_{27}^+$ ; RDA cleavage), 261 ( $[M+H-268+2H]^+$ ,  $C_{19}H_{33}^+$ ; C(13)–C(18) and C(14)–C(15) cleavage), and 289 ( $[M+H-140]^+$ ,  $C_{20}H_{33}O^+$ ; C(6)–C(7) and C(9)–C(10) cleavage). Further HMBCs were also observed between the H-atom signal at  $\delta(H)$  2.21 (H–C(18)) and C-atom signals at  $\delta(C)$  142.2 (C(13)), 121.6 (C(12)), 34.6 (C(17)), and 37.4 (C(22)); and between H-atom signal at  $\delta(H)$  1.83–1.81 ( $H_a$ –C(22)) and C-atom signals at  $\delta(C)$  32.7 (C(21)) and 20.0 (C(28); Fig. 2). A NOESY correlation between H-atoms at  $\delta(H)$  3.40–3.33 (H–C(16)) and 0.86 (Me(26), Fig. 3) suggested the  $\alpha$ -configuration of the OH group at position C(16). On the basis of the NMR data above and by comparison with reported lupane-type triterpenoids

[12][14][15], the structure of clerodendrumol (1) was elucidated as (16 $\alpha$ )-lupa-12,20(29)-dien-16-ol (1).

**Antiproliferative Activity.** The antiproliferative activities of the crude extract, compounds 1, 2, 3, and 5 were determined on five cancer cell lines (Table 2). The human breast cancer cell line (MDA-MB-231) was sensitive to all test samples. Clerodendrumol (1) and its derivative (2) had good activity ( $IC_{50}$  values 7.5 and 13.2  $\mu M$ ) on this cell line, while the crude extract ( $IC_{50}$  39.2  $\mu g/ml$ ), and compounds 3 ( $IC_{50}$  24.7  $\mu M$ ) and 5 ( $IC_{50}$  38.4  $\mu M$ ) were only moderately active. Genkwanin (5) exhibited strong activity on HeLa cell line with  $IC_{50}$  (7.8  $\mu M$ ) comparable to that of the reference standard 5-fluorouracil (6.4  $\mu M$ ), whereas the crude extract had only weak activity ( $IC_{50}$  92.6  $\mu g/ml$ ). The antiproliferative activity of genkwanin (5) is in agreement with the previous result obtained for a flavonoid on HeLa cancer cell line [16].

Lupane-type triterpenoids have been reported to possess antiproliferative activity on cancer cell lines [17–19]. Thus, our results on the isolation of an antiproliferative triterpenoid, clerodendrumol (1), from *Clerodendrum yaundense* strengthen the previous findings. The phytochemical study of others species of the same genus is in progress in our research group, as well as the characterization of more bioactive constituents.

### Experimental Part

**General.** TLC: Pre-coated silica gel  $GF_{254}$  plates ( $SiO_2$ ; Merck) with detection accomplished by visualizing with a UV light (254 and 365 nm), followed by spraying with 1%  $Ce(SO_4)_2/10\%$  aq.  $H_2SO_4$  and then heating to 150°. Column chromatography (CC): silica gel 60N

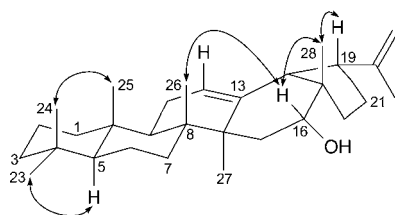


Fig. 3. Key NOESY (H ↔ H) of clerodendrumol (1)

Table 1.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Data (500 and 125 MHz, resp.) for *Clerodendrum* (**1**) and *Acetylclerodendrum* (**2**).  $\delta$  in ppm,  $J$  in Hz.

| Position | <b>1<sup>a</sup></b>  |                    | <b>2<sup>b</sup></b>                              |                    |
|----------|---|--------------------|---|--------------------|
|          | $\delta(\text{H})$  | $\delta(\text{C})$ | $\delta(\text{H})$                                | $\delta(\text{C})$ |
| 1        | 1.95–1.93 ( <i>m</i> ), 1.99–1.97 ( <i>m</i> )                              | 32.6               | 1.50–1.48 ( <i>m</i> ), 2.00–1.98 ( <i>m</i> )    | 33.9               |
| 2        | 1.53–1.51 ( <i>m</i> ), 1.55–1.53 ( <i>m</i> )                              | 20.2               | 1.07–1.05 ( <i>m</i> ), 1.59–1.57 ( <i>m</i> )    | 21.5               |
| 3        | 1.76–1.74 ( <i>m</i> ), 1.79–1.77 ( <i>m</i> )                              | 32.9               | 0.95–0.93 ( <i>m</i> ), 1.00–0.98 ( <i>m</i> )    | 33.7               |
| 4        |   | 36.4               |   | 36.2               |
| 5        | 1.02 ( <i>br. s</i> )   | 57.7               | 1.01–0.99 ( <i>m</i> )                            | 56.7               |
| 6        | 1.58–1.56 ( <i>m</i> ), 1.71–1.69 ( <i>m</i> )                              | 23.8               | 1.17–1.15 ( <i>m</i> ), 1.34–1.32 ( <i>m</i> )    | 24.3               |
| 7        | 1.30–1.28 ( <i>m</i> ), 2.03–2.01 ( <i>m</i> )                              | 29.0               | 0.98–0.96 ( <i>m</i> ), 1.32–1.30 ( <i>m</i> )    | 33.9               |
| 8        |   | 51.3               |   | 56.7               |
| 9        | 1.08 ( <i>br. s</i> )   | 57.0               | 0.94–0.92 ( <i>m</i> )                            | 50.0               |
| 10       |   | 46.8               |   | 42.3               |
| 11       | 1.47 ( <i>dd</i> , $J = 5.7, 3.5$ ),<br>1.43 ( <i>dd</i> , $J = 6.3, 4.6$ ) | 38.3               | 1.14–1.12 ( <i>m</i> ),<br>1.86–1.84 ( <i>m</i> ) | 37.0               |
| 12       | 5.31 ( <i>br. d</i> , $J = 4.6$ )   | 121.6              | 5.37 ( <i>d</i> , $J = 2.7$ )                     | 122.7              |
| 13       |   | 142.2              |   | 139.7              |
| 14       |   | 43.2               |   | 45.8               |
| 15       | 2.03 ( <i>br. d</i> , $J = 3.4$ ),<br>1.98 ( <i>dd</i> , $J = 5.7, 3.4$ )   | 40.8               | 2.32–2.30 ( <i>m</i> ),<br>2.33–2.31 ( <i>m</i> ) | 38.1               |
| 16       | 3.40–3.33 ( <i>m</i> )  | 71.8               | 4.60 ( <i>t</i> , $J = 4.9$ )                     | 74.0               |
| 17       |   | 34.6               |   | 36.6               |
| 18       | 2.21 ( <i>d</i> , $J = 5.7$ )   | 43.4               | 1.60 ( <i>br. s</i> )                             | 31.9               |
| 19       | 1.88–1.86 ( <i>m</i> )  | 50.4               | 1.86 ( <i>d</i> , $J = 10.9$ )                    | 49.5               |
| 20       |   | 148.2              |   | 147.6              |
| 21       | 1.04–1.03 ( <i>m</i> ), 1.06–1.05 ( <i>m</i> )                              | 32.7               | 1.57–1.55 ( <i>m</i> ), 1.88–1.86 ( <i>m</i> )    | 27.8               |
| 22       | 1.83–1.81 ( <i>m</i> ), 1.86–1.84 ( <i>m</i> )                              | 37.4               | 1.51–1.49 ( <i>m</i> ), 1.96–1.94 ( <i>m</i> )    | 39.7               |
| 23       | 0.73 ( <i>s</i> )   | 34.6               | 1.25 ( <i>s</i> )                                 | 35.5               |
| 24       | 0.95 ( <i>s</i> )   | 22.0               | 0.91 ( <i>s</i> )                                 | 21.5               |
| 25       | 0.72 ( <i>s</i> )   | 19.3               | 0.84 ( <i>s</i> )                                 | 19.8               |
| 26       | 0.86 ( <i>s</i> )   | 12.5               | 0.67 ( <i>s</i> )                                 | 12.0               |
| 27       | 1.02 ( <i>s</i> )   | 26.8               | 1.02 ( <i>s</i> )                                 | 29.4               |
| 28       | 1.02 ( <i>s</i> )   | 20.0               | 1.02 ( <i>s</i> )                                 | 19.3               |
| 29       | 4.74 ( <i>br. s</i> ), 4.67 ( <i>br. s</i> )                                | 112.2              | 4.73 ( <i>br. s</i> ), 4.64 ( <i>br. s</i> )      | 111.4              |
| 30       | 1.58 ( <i>s</i> )   | 18.0               | 1.60 ( <i>s</i> )                                 | 17.8               |
| MeC=O    | –   | –                  | –   | 170.6              |
| MeC=O    | –   | –                  | 2.03 ( <i>s</i> )                                 | 21.0               |

<sup>a</sup>) Recorded in ( $\text{D}_6$ )acetone. <sup>b</sup>) Recorded in  $\text{CDCl}_3$ .

Table 2. Antiproliferative Activity of Crude Extract and Compounds<sup>a</sup>) Isolated from *C. yaundense*

| Samples                       | $IC_{50}$ |            |       |       |        |
|-------------------------------|-----------|------------|-------|-------|--------|
|                               | HeLa      | MDA-MB 231 | PSN-1 | A549  | PANC-1 |
| Crude extract <sup>b</sup> )  | 92.6      | 39.2       | > 100 | > 100 | 88.5   |
| <b>1</b> <sup>c</sup> )       | > 100     | 7.5        | > 100 | > 100 | > 100  |
| <b>2</b>                      | > 100     | 13.2       | > 100 | > 100 | > 100  |
| <b>3</b>                      | > 100     | 24.7       | > 100 | > 100 | > 100  |
| <b>5</b>                      | 7.8       | 38.4       | 88.9  | > 100 | 49.3   |
| 5-Fluorouracil <sup>d</sup> ) | 6.4       | 1.1        | 8.7   | 9.7   | 3.0    |

<sup>a</sup>) Compounds **4**, **6**, and **7** were not tested. <sup>b</sup>)  $\mu\text{g/ml}$ . <sup>c</sup>)  $IC_{50}$  in  $\mu\text{M}$ . <sup>d</sup>) Positive control,  $IC_{50}$  in  $\mu\text{M}$ .

spherical, neutral, 40–50  $\mu\text{m}$  (*Kanto*, Japan), *Cosmosil 75C<sub>18</sub>*-silica gel (*Kyoto*, Japan), and *Sephadex LH-20*. MPLC: *Büchi Sepacore* system (*Büchi Labortechnik AG*, Flawil, Switzerland). M.p.: *Yanaco, Micro MP* apparatus; uncorrected. Optical rotations: *Jasco P-2100* polarimeter. IR Spectra: *Jasco FT-IR 460* spectrometer (*Kyoto*, Japan);  $\tilde{\nu}$  in  $\text{cm}^{-1}$ . NMR Spectra: *Bruker 500* instrument;  $\delta$  in ppm rel. to  $\text{Me}_4\text{Si}$  as internal standard,  $J$  in Hz. FAB-MS: *Jeol JMS-AX505W* spectrometer; in  $m/z$ .

**Plant Material.** The twigs of *Clerodendrum yaundense* were collected in Dschang (Western-Region of Cameroon) in April 2015 and were identified by Mr. *Nana*, a retired botanist, from the Yaoundé National Herbarium where our sample was compared to the available specimen with voucher number 10842 SRF/Cam.

**Extraction and Isolation.** The twigs of *C. yaundense* (2.8 kg) were dried and milled to a powder before macerated in MeOH (10 l) for 24 h (repeated three times) to give a crude extract (102 g, 4% yield) after

filtration and removal of the solvent using rotative evaporator. Part of this extract (50 g) was subjected to a normal phase SiO<sub>2</sub> MPLC using hexane, AcOEt, and MeOH as eluent in gradient polarity to give 73 fractions of 300 ml each that were monitored by TLC to ten combined fractions (Fr. F<sub>1</sub>–F<sub>10</sub>). Fr. F<sub>2</sub> (500 mg, hexane/AcOEt 95:5) gave **4** (19 mg), while Fr. F<sub>3</sub> (900 mg, hexane/AcOEt 9:1) crystallized in acetone to give a mixture of **1** and **3** (197 mg). After unsuccessful separation of this mixture using *Sephadex LH-20* and silica gel reversed and normal phases column chromatography (CC), chemical reactions (acetylation and deacetylation) were used to finally obtain compounds **1**, **2**, and **3** in pure form. Fr. F<sub>6</sub> (240 mg, hexane/AcOEt 7:3) was subjected to the reversed phase SiO<sub>2</sub> CC using MeOH and H<sub>2</sub>O in gradient polarity to give 74 fractions of 50 ml each that were further combined by comparative TLC to 17 fractions. Frs. F<sub>6-3</sub>, F<sub>6-6</sub>, and F<sub>6-8</sub> were separately subjected to prep. TLC followed by *Sephadex LH-20* to afford compounds **5** (34.8 mg), **6** (8.3 mg), and **7** (2.4 mg), resp.

**Acetylation of the Mixture of 1 and 3.** Acetic anhydride (2 ml) was added to a mixture of compounds **1** and **3** (100 mg) dissolved in pyridine (2 ml) and kept in the darkness at r.t. for 24 h. The reaction was quenched with 10% HCl and extracted with AcOEt to yield **1** (64.1 mg, 64%) and **2** (30.4 mg, 31%) after purification on SiO<sub>2</sub> CC using hexane/AcOEt (4:1).

**Deacetylation of 2.** Compound **2** (50 mg) was dissolved in CHCl<sub>3</sub>/MeOH 1:9 (10 ml), and a 2M soln. of K<sub>2</sub>CO<sub>3</sub> (10 ml) was added. After stirring at r.t. for 4 h, the TLC (hexane/AcOEt 4:1) showed complete consumption of **2** with one different and more polar product at R<sub>f</sub> 0.3. Neutralization with a sat. soln. of NH<sub>4</sub>Cl (15 ml) followed by extraction using AcOEt and removal of residual H<sub>2</sub>O by anh. Na<sub>2</sub>SO<sub>4</sub> gave compound **1** (43.5 mg, 87% yield).

**Clerodendrumol** (= (16 $\alpha$ )-Lupa-12,20(29)-dien-16-ol; **1**). White powder. M.p. 232–234°. [ $\alpha$ ]<sub>D</sub><sup>25</sup> = –75.3 (c = 0.1, acetone). IR: 3438, 2938, 1645, 1465, 1377, 1053, 886. <sup>1</sup>H- (500 MHz, (D<sub>6</sub>)acetone) and <sup>13</sup>C-NMR (125 MHz, (D<sub>6</sub>)acetone): see Table 1. FAB-MS: 425 (18), 411 (3), 395 (5), 307 (18), 289 (5), 261 (3), 217 (22), 195 (4), 154 (100), 136 (77). HR-FAB-MS: 425.3781 ([M + H]<sup>+</sup>, C<sub>30</sub>H<sub>49</sub>O<sup>+</sup>; calc. 425.3784).

**O-Acetylclerodendrumol** (= (16 $\alpha$ )-Lupa-12,20(29)-dien-16-yl Acetate; **2**). White powder. M.p. 215–217°. [ $\alpha$ ]<sub>D</sub><sup>25</sup> = –21.7 (c = 0.1, acetone). IR: 2937, 1732, 1646, 1467, 1368, 1251, 1039, 886. <sup>1</sup>H- (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>): see Table 1. FAB-MS: 467 (8), 453 (25), 395 (30), 255 (10), 159 (20), 105 (80), 81 (100). HR-FAB-MS: 467.38833 ([M + H]<sup>+</sup>, C<sub>30</sub>H<sub>51</sub>O<sub>2</sub><sup>+</sup>; calc. 467.38891).

**In vitro Antiproliferative Assay.** The cell lines used were A549 (human lung cancer), HeLa (human cervix cancer), PANC-1 and PSN-1 (human pancreatic cancer), and MDA-MB-231 (human breast cancer).  $\alpha$ -Minimum essential medium with L-glutamine and phenol red ( $\alpha$ -MEM, Wako) was used for the first three cell lines, whereas high-glucose Dulbecco's modified Eagle's medium with L-glutamine, phenol red, and sodium pyruvate (DMEM, Wako) were used for the latter ones. Both media were supplemented with 10% fetal bovine serum (*Nichirei Bioscience*) and 1% antibiotic antimycotic soln. (*Sigma–Aldrich*). The *in vitro* antiproliferative activity of the crude extract and isolated compounds was determined as described previously [20]. Briefly, each cell line was seeded in 96-well plates (2 × 10<sup>3</sup> per well) and incubated at 37° for 24 h either in  $\alpha$ -MEM or in DMEM under 5% CO<sub>2</sub> and 95% air. Thereafter, cells were washed with PBS (*Nissui Pharmaceuticals*), and test samples in serial dilution were added. After 72 h incubation, cells were washed with PBS, and 100  $\mu$ l of  $\alpha$ -MEM or DMEM containing 10% WST-8 cell counting kit (*Kumamoto, Japan*) sol. was added to the wells. The absorbance was measured at 450 nm after additional 2 h incubation.

The different concentrations of serial dilutions of tested samples were 100–3.125  $\mu$ g/ml for the crude extract, 100–3.125  $\mu$ M for isolated compounds, and 10–0.3125  $\mu$ M for the positive control (5-fluorouracil), resp. Cell viability was calculated from the mean values of data from three wells by using the following equation, and antiproliferative activity was expressed as IC<sub>50</sub> (50% inhibitory concentration) value.

$$\% \text{ Cell viability} = 100 \times (\text{Abs}_{\text{sample}} - \text{Abs}_{\text{blank}}) / (\text{Abs}_{\text{control}} - \text{Abs}_{\text{blank}})$$

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